(FILE 'HOME' ENTERED AT 15:56:11 ON 26 JUL 2007)

FILE 'CA' ENTERED AT 15:56:26 ON 26 JUL 2007

- L1 3243 S (FLUORESCEN? OR FLUORESCING OR LUMINESCEN? OR LUMINESCING) AND (NON(W) (FRET OR FLUORESCEN?) OR NONFRET OR NONFLUORESC?)
- L2 605 S L1 AND (QUENCH? OR DIMER OR CLOSE(1A)CONTACT? OR DYE(1W)DYE OR HYDROPHOBIC(2A)INTERACT?)
- 106 S L1 AND (QUENCH? OR DIMER OR DILABEL? OR (DUAL OR DI OR DOUBLE OR BI OR TWO OR PAIR) (1A) (FLUOROPHORE OR CHROMOPHORE OR DYE OR INDICATOR OR LABEL?) OR BILABEL? OR DONOR OR ACCEPTOR) AND (STATIC OR GROUND STATE OR STACK? OR CLOSE (1A) CONTACT? OR DYE (1W) DYE OR HYDROPHOBIC (2A) INTERACT?)
- L4 34 S L2 AND (PEPTIDE OR POLYPEPTIDE OR POLYAMINO? OR POLY AMINO?)
- L5 33 S L1 AND KINASE
- L6 324 S L1 AND(DIMER OR DILABEL? OR (DUAL OR DI OR DOUBLE OR BI OR TWO OR PAIR) (1A) (FLUOROPHORE OR CHROMOPHORE OR DYE OR INDICATOR OR LABEL?) OR BILABEL? OR DONOR OR ACCEPTOR)
- L7 82 S L6 AND (PEPTIDE OR POLYPEPTIDE OR PROTEIN OR ENZYM? OR POLYAMINO?)
- L8 71 S L1 AND QUENCH?(6A)(DIMER OR DILABEL? OR (DUAL OR DI OR DOUBLE OR BI OR TWO OR PAIR)(1A) (FLUOROPHORE OR CHROMOPHORE OR DYE OR INDICATOR OR LABEL?) OR BILABEL? OR DONOR OR ACCEPTOR OR STATIC OR GROUND STATE OR STACK? OR CLOSE(1A)CONTACT? OR DYE(1W)DYE OR HYDROPHOBIC(2A)INTERACT?)
- L9 15 S L1 AND (EXIMER OR EXCIMER OR EXIPLEX OR EXCIPLEX) (6A) QUENCH?
- L10 257 S L3-5, L7-9
- L11 140 S L10 AND PY<2000
- L12 20 S L10 AND PATENT/DT

FILE 'BIOSIS' ENTERED AT 16:34:36 ON 26 JUL 2007

L13 67 S L11

FILE 'MEDLINE' ENTERED AT 16:35:22 ON 26 JUL 2007

- L14 62 S L11
 - FILE 'CA, BIOSIS, MEDLINE' ENTERED AT 16:36:29 ON 26 JUL 2007
- L15 175 DUP REM L11 L12 L13 L14 (114 DUPLICATES REMOVED)
- => d bib, ab 115 1-175
- L15 ANSWER 33 OF 175 CA COPYRIGHT 2007 ACS on STN
- AN 128:252109 CA
- TI Limitations of **quenching** as a method of fluorometric analysis of **non-fluorescent** analytes
- AU Rakicioglu, Yener; Mickey Young, Melissa; Schulman, Stephen G.
- CS Gainesville, College of Pharmacy, University of Florida, FL, 32610, USA
- SO Analytica Chimica Acta (1998), 359(3), 269-274
- Diffusional quenching of ordinary fluorophores (with decay times no longer than several tens of nanoseconds), a process employed in many fluorescence optical sensors, is rather insensitive as a anal. method having limits of detection no better than \$\Omega\$10-4 M. Longer lived luminophores, such as lanthanides, actinides and phosphorophores, however, can be quenched with substantially lower detection limits for the quencher. Static quenching, however, is limited by the thermodn. strength of the complex(es) formed between the fluorophore and quencher

rather than by the decay time of the fluorophore and the diffusibility of the **quencher** and can be quite sensitive as an anal. method for the **quencher**.

- L15 ANSWER 155 OF 175 CA COPYRIGHT 2007 ACS on STN
- AN 83:110396 CA
- TI Long-range quenched peptides as fluorogenic substrates of proteolytic enzymes
- AU Carmel, Amos
- CS Dep. Biophys., Weizmann Inst. Sci., Rehovot, Israel
- SO Pept., Proc. Eur. Pept. Symp., 13th (1975), Meeting Date 1974, 385-91. Editor(s): Wolman, Yecheskel. Publisher: Wiley, New York, N. Y.
- AΒ New techniques were developed for monitoring the hydrolysis of peptides by proteinases by using the principle that the emissive features of a fluorescent chromophore are affected not only by the immediate environment but also by interactions with moieties which are located at some distance, either in soln. or on the same mol. backbone. Modifications in the geometry of such a system can therefore be followed by a change in the extent of these interactions as reflected in the fluorescence spectrum. In a compd. such as anthracene 9-carbonyl- β alanyllysylalanyl-2-naphthylmethylamide-HBr (I), the **donor** fluorophore is the naphthalene moiety while the acceptor fluorophore is the anthracene moiety. In such a compd. there is an overlap between the emission spectrum of the donor and the absorption spectrum of the acceptor which can be expressed quant. as an integral equation. method was applied to the study of the tryptic digestion of I and 3 homologous oligolysines blocked by the same chomophores, anthracene 9carbonyl- β -alanyl-(Lys)2-4-2-naphthylmethylamide. The p-nitrobenzyl ester-2HBr's of the **peptides** ϵ -anthranilyllysylalanine, ϵ anthranilyllysylphenylalanine, and ε -anthranilyllysylalanylalanine were similarly studied with leucine aminopeptidase. The p-nitrobenzyl ester-HBr and -2HBr, resp., of anthracene 9-carbonyl- β -alanyllysylalanine and tryptophyllysylalanine were hydrolyzed by porcine elastase and pnitrophenylalanylalanyllysyl-1-naphthylaminoethylamide-3HBr was a substrate for Clostridium histolyticum aminopeptidase. The new techniques possess these advantages. Suitable pairs of either donoracceptor or fluorophore-p-nitrobenzyl (nonfluorescent) groups can be introduced into the substrate mol. at chosen intervals so as not to participate directly in the enzymic reaction. Hence, changes in fluorescence can reflect directly the rate of cleavage of the susceptible bonds. Another advantage is the great versatility in prepg. potential substrates.

^{=&}gt; log y STN INTERNATIONAL LOGOFF AT 16:37:39 ON 26 JUL 2007